Cellulose trinitrate: molecular conformation and packing considerations

D. Meader, E. D. T. Atkins and F. Happey*

H. H. Wills Physics Laboratory, University of Bristol, Royal Fort, Tyndall Avenue, Bristol BS8 1TL, UK (Received I August 1978)

X-ray diffraction patterns from highly oriented fibres of cellulose trinitrate indicate that the molecule crystallizes in a five-fold helical conformation with an axial advance per monomer of 0.508 nm. Computerized molecular model building studies favour a 52 helix, i.e. a right-handed helix with two complete turns of the backbone in the layer line repeat of 2.54 nm. Two possible unit cells, one of which has been proposed previously (Happey, *F. J. Text. Inst.* 1950, 41,381), are described and the packing of the chains is discussed.

INTRODUCTION EXPERIMENTAL

Analyses of the X-ray diffraction patterns obtained from *Sample preparation* native cellulose fibres argue for crystalline domains of chains running parallel to the fibre axis and with a common pola-

running parallel to the fibre axis and with a common pola-

rity^{1,2} Fach chain has a two-fold helical conformation with acid/acetic acid/acetic anhydride or ni rity^{1,2}. Each chain has a two-fold helical conformation with acid/acetic acid/acetic anhydride or nitric acid/phosphoric
an avial advance ner p glucose monomer of 0.52 nm. The acid/phosphoric anhydride mixtures which ca an axial advance per D-glucose monomer of 0.52 nm. The acid/phosphoric annydride mixtures which caused the $-OH$
natterns exhibit layer lines with a spacing of twice this value groups in the cellulose to be replaced by patterns exhibit layer lines with a spacing of twice this value at 1.04 nm.

immersing the native fibres in aqueous solutions of mixtures of nitric, acetic and phosphoric acid, provide a series of of more, above and prospheric acts, provide a series of *X-ray diffraction*
quite different X-ray diffraction patterns³. The maximum *X-ray diffraction* photographs were recorded on a flat
X-ray diffraction photographs theoretical nitrogen content, corresponding to cellulose X-ray diffraction photographs were recorded on a flat
trinitrate, is 14.14%. In practice this is never achieved, the film, fibre camera using Ni-filtered CuKa radiat trinitrate, is 14.14%. In practice this is never achieved, the film, fibre camera using Ni-filtered CuKa radiation and pin-
nitration proceeding until an equilibrium is reached, typi-
hole collimation. The speciment to fil nitration proceeding until an equilibrium is reached, typically somewhere between the dinitrate and trinitrate

derivatives. *Molecular model building Figure 1* illustrates a typical fibre-type X-ray diffraction Molecular models were generated using a linked-atom
pattern from highly nitrated cellulose (13.9% N) and native procedure. An outline of the method has been rep pattern from highly nitrated cellulose (13.9% N) and native procedure. An outline of the method has been reported cellulose for comparison. The nitrated derivative exhibits $\frac{1}{2}$ by Gardner et al.⁵ while a more rigo cellulose for comparison. The nitrated derivative exhibits by Gardner *et al.*⁵, while a more rigorous description has layer lines of spacing 2.54 nm which is five times the expec-
heen given by Smith and Arnott⁶ Stere layer lines of spacing 2.54 nm which is five times the expec-
ted projection of a single monomer. A meridional reflection[†] meters for the cellulose backbone were taken from an a ted projection of a single monomer. A meridional reflection meters for the cellulose backbone were taken from an ave-
is observed on the fifth layer line (spacing 0.508 nm) which rage set derived by Arnott and Soott⁷ fo suggests some type of five-fold helix for the highly nitrated due in the 4C_1 chair conformation cellulose chain, with an axial advance per monomer (h) Very little information is available. cellulose chain, with an axial advance per monomer (h) Very little information is available about the detailed
slightly less than the value observed for the native cellulose structures of nitrate esters. Early work⁸ in slightly less than the value observed for the native cellulose structures of nitrate esters. Early work⁸ indicated a non-
chain.

In this contribution we have scrutinized the X-ray diffrac-
tion results in more detail and, using computerized model-
A mean set of values for the starsochanical parameters of tion results in more detail and, using computerized model-
building procedures, have investigated the stereochemical
the coNO₂ group is shown in *Figure 2a*. Because insuffibuilding procedures, have investigated the stereochemical the -ONO₂ group is shown in *Figure 2a*. Because insuffi-
feasibility of a number of models exhibiting five-fold helical cient evidence could be found to assign v feasibility of a number of models exhibiting five-fold helical cient evidence could be found to assign values to the
symmetry. In addition we have briefly considered the pack-
 $C = D - N$ bond angles β_1 , β_2 and β_2 symmetry. In addition we have briefly considered the pack-
ing of such helices in several unit cells especially with refe-
taken as variables in the model building procedure and a ing of such helices in several unit cells especially with refe-

rence to previous suggestions by Mathieu⁴ and Happey³.

weighting scheme was employed to restrict their allowed

Nitrated derivatives of cellulose, typically prepared by acid, samples with a nitrogen content of between 11% and
mersing the native fibres in aguacus solutions of mixtures 13.9% were produced.

rage set derived by Arnott and Scott⁷ for the pyranose resi-

ain.
In this contribution we have scrutinized the X-ray diffrac-
 $\frac{\text{planar geometry}}{\text{cent studies}}$ strated the $\frac{-\text{ONO}_2}{\text{syn studies}}$ group; however, more rerence to previous suggestions by Mathieu and Happey 3. weighting scheme was employed to restrict their allowed values to the range $109.5^{\circ}-120^{\circ}$. Apart from this, all bond * Permanent address: Whirlow, Lon Refail, Llanfairpwll, Anglesey, lengths and bond angles were held constant, including the discover of the constant of the constant of the set of the constant of the constant of the set of Emeritus Professor, University of Bradford.
Some weaker meridional arcs are often observed on other layer
Torsion angles τ_t , to τ_0 and bond angles β_t to β_0 were a

Torsion angles τ_1 to τ_9 and bond angles β_1 to β_3 were al-

lines which do not readily relate to the expected monomer repeat lines which do not readily relate to the expected monomer repeat
distance and are thought to arise from the slight non-stoichiometry lowed to vary subject to constraints imposing helical symof the nitrate groups.

 $(13.9\%$ nitrogen) showing a layer line spacing of 2.54 nm and exhibiting five-fold helical symmetry; (b) native cellulose showing a $N(6)$ $N(5)$

defined by these constraints the number of close intrachain b $O(4)$ I contacts between atoms was minimized.

and $5₄$ were investigated^{\ddagger}. In this notation the subscript refers to the number of complete turns of the helix in the angles and variable bond angles are marked τ_1 to τ_9 and β_1 to β_3 , $axial repeat distance (2.54 nm).$ respectively

RESULTS AND DISCUSSION

Conformation of a single chain

Sathyanarayana and Rao¹² have shown that the $5₁$ and 54 conformations are impossible for a molecule with a $(1 \rightarrow 4)$ -linked β -D-glucose backbone and with a value of h

= 0.508 nm. Our calculations confirm this conclusion.
Although it was possible to build $5₃$ helices, these invariably gave unacceptable close contacts between some

respectively, the second burst control of the parameters given by Scott and Scheraga ¹³

atoms in the chain backbone. Only in the 52 helical confora mation could reasonably satisfactory models be constructed. At this level of analysis it was not possible to distinguish between models with 0(6) near the *gg, gt* and *tg* positions § as in each case the conformation of the backbone and the sidegroups on $C(2)$ and $C(3)$ were identical, and the side-group on $C(6)$ did not give rise to any over-short contacts. However, each of these models incurs two short contacts as given in *Table 1*. Since these contacts are not severe, we would anticipate that they could be removed by slight variations in the glycopyranose ring geometry. The Cartesian coordinates of the asymmetric unit of the most acceptable models are listed in *Table 2* and the three side-group orientations indicated. Computer-drawn projections of the *gt* case are shown in *Figure 3.*

Unit cell considerations

The measured d spacings taken from the X-ray diffraction pattern shown in *Figure la* are listed in *Table 3.*

Mathieu⁴ originally proposed a unit cell with the dimen**b** sions: $a = 0.90$ nm, $b = 1.39$ nm, c (fibre axis) = 2.56 nm, α $= \beta = \gamma = 90^\circ$. Happey³ argued that Mathieu's cell was incorrectly calculated from the crystallographic data and pos-

§ Pure *gg* is $\tau_7 = 60^\circ$, *gt* is $\tau_7 = 180^\circ$ and *tg* is $\tau_7 = -60^\circ$

The four possible five-fold helical symmetries $51, 52, 53$ *Figure 2* (a) Mean bond distances and valency angles for the
d 5. were investigated $\ddot{\textbf{i}}$ In this notation the subscript $O-NO_2$ group^{9,10,11}. (b) Schema monomer with appropriate atoms labelled. The nine torsional

Table 1 Close intrachain contacts for the most acceptable $5₂$ helical conformations

	Separation (nm) Extreme	Contact criteria ¹⁴ (nm)		
Atoms			Normal	(kJ/mol)
$O(3) - H'(1)$ $N(3) - C(2)$	0.214 0.270	0.220 0.280	0.240 0.290	5.8 2.1

The prime refers to the next residue in the chain. * The energy was $^+$ The 53 and 54 helices are left-handed versions of 5₂ and 5₁ calculated using a standard Lennard-Jones potential function with

Unit common to all models

Atom	X (nm)	Y (nm)	Z (nm)	
O(1)	0.1064	0.0000	0.0000	
C(1)	0.0352	-0.0145	0.1184	
C(2)	0.1317	-0.0590	0.2274	
C(3)	0.0618	-0.0625	0.3624	
C(4)	-0.0056	0.0711	0.3906	
C(5)	-0.0947	0.1109	0.2734	
O(5)	-0.0182	0.1138	0.1519	
H(1)	-0.0469	-0.0873	0.1100	
O(2)	0.1843	-0.1870	0.1944	
H(2)	0.2167	0.0108	0.2319	
O(3)	0.1570	-0.0911	0.4651	
H(3)	-0.0138	-0.1424	0.3625	
O(4)	-0.0861	0.0625	0.5080	
H(4)	0.0712	0.1484	0.4062	
C(6)	-0.1556	0.2484	0.2903	
H(5)	-0.1768	0.0383	0.2634	
N(2)	0.1042	-0.2962	0.2163	
O(2A)	0.0206	-0.3238	0.1319	
O(2B)	0.1209	-0.3597	0.3191	
N(3)	0.2839	-0.0415	0.4495	
O(3A)	0.3014	0.0778	0.4682	
O(3B)	0.3723	-0.1193	0.4178	

0(6) nearly *gg*

Atom	X (nm)	Y (nm)	Z (nm)	⊀
H(6A)	-0.0755	0.3231	0.3003	
H(6B)	-0.2185	0.2499	0.3806	
O(6)	-0.2375	0.2849	0.1776	
N(6)	-0.3707	0.2776	0.2091	
O(6A)	-0.4254	0.1688	0.2021	
O(6B)	-0.4274	0.3804	0.2425	

The helix axis corresponds to the Z-axis of the Cartesian coordinate system used

tulated a modified cell, assuming the same indexing. This corrected cell has the b -axis contracted to 1.225 nm with the a -axis and angles remaining unaltered. The c -axis takes the slightly smaller value of 2.54 nm. The calculated d spacings from this modified cell are also given in *Table 3.* While rescrutinising the measured d spacings it was noticed that if Mathieu's indexing was not assumed an alternative unit cell is possible with dimensions: $a = 0.90$ nm, $b =$ 1.46 nm, c(fibre axis) = 2.54 nm. The calculated d spacings *Figure 3* Projections of the proposed 5₂ helix for cellulose trinitrate for this cell are also given in *Table 3.* The paucity of obser- with 0(6) *gt*

Table 2 Cartesian coordinates of the asymmetric repeating unit for ved reflections in the X-ray diffraction pattern make it diffitible most acceptable 5₂ helical conformations cult to define a unique cell on this information alone.

$Packing$ *considerations*

Hexagonal close packing is the most symmetrical and commonly observed arrangement of polymeric molecules having approximately circular cross-section; each molecule is equidistant from its six nearest neighbours. Clearly the five-fold nature of the cellulose trinitrate molecule perturbs this packing arrangement somewhat, giving rise to additional Bragg reflections and larger unit cells. The lattice with $a =$ 0.90 nm, $b = 1.46$ nm has the advantage that the angle between the diagonals is close to 60° (63.3°). If we place molecules at the corners and centre of such a cell then the chains pack approximately on a hexagonal lattice. This arrangement, which we feel is representative of the essential features of the juxtapositioning of the molecules, is illustrated in *Figure 4*.

It must be emphasised that the proposed models were built in isolation. It is not surprising, therefore, that some over-short interchain contacts arise when the chains are placed on the lattice sites as in *Figure 4*. Owing to the difficulties in defining a unique unit cell, we can draw no firm

Cellulose trinitrate: molecular conformation and packing considerations: D. Meader et aL

Figure 4 Projection of tentative lattice for cellulose trinitrate with 8 ⁵Structure Reports', 1942–1944, 9, 313
O(6) gt. The projections down the chain axis are oriented randomly 9 Dixon, W. B. and Wilson, E. B. J. Che O(6) *gt.* The projections down the chain axis are oriented randomly 9 on a face centred lattice with $a = 0.90$ nm and $b = 1.46$ nm. This is 10 on a face centred lattice with a = 0.90 nm and b = 1.46 nm. This is 10 Trotter, J. Acta Crystallogr. 1963, 16, 698
approximately a hexagonal array. Some chains appear with opposite 11 Yukawa, Y. 'Handbook of Organic Struct approximately a hexagonal array. Some chains appear with opposite 11
polarity

conclusions as to the most likely polarity of the chains but $\frac{10,1605}{2}$ Scott, R. A. and Scheraga, H. A. J. Chem. Phys. 1966, 45, a crude analysis indicates that the proposed models have most room when they are antiparallel. For this reason some 14 of the chains in *Figure 4* are shown inverted. 23, 283

$ACKNOWLEDGEMENTS$

We thank the Science Research Council for support, including a studentship to D. M. One of the authors (F. H.) wishes to thank the Leverhulme Research Trust for an Emeritus Fellowship.

REFERENCES

- 1 Gardner, K. H. and Blackwell, J. *Biopolymers* 1974, 13, 1975
- 2 Sarko, A. and Muggli, R. *Macromolecules* t974, 7,486
- }~ .~.~ ~ ~ ~ ~f'~ ~k 3 Happey, F.J. *Text. Inst.* 1950,41,381
-
- 4 Mathieu, *M. C. R. Acad. Sci. (Paris)* 1935,200, 401 5 Gardner, K. H., Magill, J. H. and Atkins, E. D. T. *Polymer* 1978, 19, 370
- 6 Smith, P. J. C. and Arnott, S. *Acta Crystallogr. (A)* 1978, 34,3
7 Arnott, S. and Scott, W. E. J. Chem. Soc. (Perkin Trans. 2).
- 7 Arnott, S. and Scott, W. E. J. *Chem. Soc. (Perkin Trans. 2),* 1972, p 324
'Structure Reports', 1942–1944, 9, 313
-
-
- -
- Benjamin, New York, 1965
12 Sathyanarayana, B. K. and 12 Sathyanarayana, B. K. and Rao, V. S. R. *Biopolymers* 1971,
-
- 2091
Ramachandran, G. N. and Sasisekharan, V. Adv. Protein Chem.